GROWTH, NUTRIENT UTILIZATION AND BODY COMPOSITION OF JUVENILE BAGRID CATFISH, *CHRYSICHTHYS NIGRODIGITATUS* (ACTINOPTERYGII: SILURIFORMES: CLAROTEIDAE), FED DIFFERENT DIETARY CRUDE PROTEIN LEVELS

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Background. The African bagrid catfish, *Chrysichthys nigrodigitatus* (Lacepède, 1803), is an omnivorous species cultured in both fresh and brackish waters because of its good growth rate, excellent taste, and high market demand. There has been little nutritional work on this economically important species. This study was designed to determine the effects of varying dietary crude protein levels on growth, nutrient utilization and body composition of juvenile *C. nigrodigitatus*.

Materials and Methods. One hundred and twenty fish with an initial mean weight of 21.33 ± 0.48 g were divided randomly among 12 tanks for triplicate groups that were fed diets containing 20%, 25%, 30%, and 35% crude protein (CP). Feed consumption and fish weight were measured biweekly for eight weeks, after which proximate composition of the fish was determined.

Results. Fish fed the 35% CP diet had the best growth performance and nutrient utilization (P < 0.05), with a mean weight gain of 88.3% ± 0.2%, feed conversion ratio (FCR) of 2.60 ± 0.12, and protein efficiency ratio (PER) of 1.10 ± 0.05 . The least weight gain ($32.1\% \pm 0.9\%$) and poorest FCR and PER (5.70 ± 0.06 and 0.87 ± 0.01 , respectively) were observed in fish fed the 20% CP diet. Body protein content was not affected by diet, but body lipid increased significantly (P < 0.05) with increasing dietary protein levels.

Conclusion. C. nigrodigitatus juveniles will grow best when fed diets containing at least 35% CP.

Keywords: bagrid catfish, Chrysichthys nigrodigitatus, dietary protein, nutrient utilization, growth

INTRODUCTION

The bagrid catfish, *Chrysichthys nigrodigitatus* (Lacepède, 1803), is an omnivorous species indigenous to Africa that is cultured in both fresh and brackish waters (Afinowi and Marioghae 1987). It has been shown to have high food conversion efficiency, with good growth and survival (Ezenwa 1982a, 1982b), as well as high quality meat and excellent taste, and hence is in great demand and highly priced (Ezenwa 1982b, Ibrahim 1988). However, little nutritional research has been carried out on this economical important species, hindering the viability of commercial production.

In determining the minimum nutrient requirements of a cultured species, protein is usually given first consideration because of its high cost and essential role for growth, tissue maintenance and reproduction. Among the catfishes, optimal dietary crude protein (CP) levels are 24%–37% for juvenile channel catfish *Ictalurus punctatus* (see: Garling and Wilson 1976; Robinson and Li 1998, 1999,

Li et al. 2000, Gibson and Gatlin 2001, Li et al. 2003, 2007, D'Abramo et al. 2008), 33%-37% for juvenile South American jundia catfish, Rhamdia quelen (see: Meyer and Fracalossi 2004), 35% for juvenile African catfish, Clarias gariepinus (see: Arowosoge 1987, Omar 1996), 40% for fingerling (< 10 g) bagrid catfish, Chrysichthys nigrodigitatus (see: Adewolu and Fagade 1996), hybrid African catfish, Clarias gariepinus × Heterobranchus bidorsalis (see: Adebayo and Quadri 2005), and European catfish, Silurus glanis (see: Bekcan et al. 2006), and 40%-42% for Asian red tail catfish, Hemibagrus (Mystus) nemurus (see: Khan et al. 1993, 1996, Ng et al. 2001). The presently reported study was undertaken to determine the effects of varying dietary CP levels on the nutrient utilization and growth of juvenile Chrysichthys nigrodigitatus, with a view to determining its protein requirement.

MATERIALS AND METHODS

Fish. Juvenile bagrid catfish, Chrysichthys nigrodigitatus,

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were collected from Asejire Lake (Oyo State, Nigeria) and transported in an ice-chest to concrete tanks located outside the Zoology Laboratory of the University of Ibadan, Nigeria. The tanks were protected from direct sunlight by a palm-leaf roof. The fish were acclimated to the experimental conditions over a 14-day period, during which they were trained to feed on pelleted diets.

Diet Formulation and Preparation. Diets with 20%, 25%, 30%, and 35% CP (designated CP20, CP25, CP30 and CP35, respectively) were formulated using Pearson's square method (Gohl 1985). The choice of feed ingredients was based on their availability, cost and protein, energy, vitamin, and mineral contents. Diet formulations and proximate compositions are presented in Table 1. Groundnut cake (9.2% CP) and fish meal (65% CP) served as the primary protein source in a ratio of 2.5 : 1, respectively, while maize (9.2% CP) and rice bran (8.2% CP) were added as the energy source in a ratio of 3 : 1, respectively. Groundnut oil served as a fatty acid and energy source while bone meal and ground oyster shell were added as mineral sources. Sodium chloride (table salt) was also added as a mineral source and for palatability. All diets were fortified with a vitamin premix to meet the requirement for catfishes (Anonymous 1976). Diet preparation was as described by Ng et al. (1998). All ingredients were finely ground, mixed with groundnut oil and some warm water, and then mixed thoroughly with a blender until a paste was formed. Pellets were made with an improvised pelleting device and dried to a constant weight in a Gallenkamp oven preset to 60°C. Dried pellets were allowed to cool and were then stored in labelled dried bottles for feeding.

Feeding trial. Concrete tanks (n = 12), each of 100-L capacity, were supplied with static dechlorinated tap water with a daily 50% exchange of water to maintain water quality and prevent ammonia build-up. A total of 120 juvenile catfish (mean weight of 21.33 ± 0.48 g SD) were divided randomly among the tanks for four triplicate dietary treatments (10 fish/tank). Food was withheld from the fish for 24 h prior to starting the experiment. The fish were then fed their respective diets at 3% of total body weight per day, as recommended by Viveen et al. (1986) for catfishes, with half the daily ration fed to the fish at 0900 h and the other half at 1800 h. Feeding was generally completed in 5–10 min and no uneaten food remained. Fish were reweighed biweekly and feeding rate was adjusted accordingly. Water quality was maintained within the levels recommended for catfish culture (Viveen et al. 1986). Temperature was 26.5 ± 0.5 °C, pH was 7.5 \pm 0.1 and dissolved oxygen was 5.0 \pm 0.5 mg \cdot $L^{-1}.$ The feeding trial lasted for 8 weeks.

Proximate Analysis. Proximate composition of the experimental diets was analyzed before commencing the experiment, while fish were analyzed at both the beginning and end of the experiment. Moisture (105°C oven, overnight), crude protein (micro-Kjeldahl method, $N \times 6.25$), crude lipid (Soxhlet extraction), ash (muffle furnace at 550°C) and crude fibre (acid/base digestion) were deter-

mined using standard procedures (Anonymous 1990). Nitrogen-free extract (NFE; carbohydrate) was calculated by taking the sum of values for crude protein, crude lipid, crude fibre, and moisture, and subtracting this from 100 (Maynard et al. 1979)

Growth and nutrient utilization parameters. Growth and nutrient utilization parameters were assessed in terms of weight gain, specific growth rate (SGR), feed conversion ratio (FCR), protein intake (PI) and protein efficiency ratio (PER), using the following formulae:

Weight gain
$$[\%] = \frac{(W_f - W_i) \times 100}{W_i}$$
, (1)

where W_f and W_i represent mean final and initial weights [g], respectively

SGR (%BW/day) =
$$\frac{(\log_e W_f - \log_e W_i) \times 100}{T},$$
 (2)

where T represents trial duration (day)

$$FCR = \frac{TFC}{(W_f - W_i)},$$
(3)

where TFC represents total amount of feed consumed [g]

$$PI [g] = \frac{TFC \times \% \text{ protein in the diet}}{100}, \qquad (4)$$

$$PER = \frac{(W_f - W_i)}{PI}$$
(5)

Statistical Analysis. Replicate values were combined after first confirming the absence of any tank effects. One way analysis of variance (ANOVA) was used to test for the effect of diet (CP level) on the various parameters measured in this study, using Microsoft STATISTICA[®]. A p-value of 0.05 was used as the level of statistical significance.

RESULTS

There was no mortality and all fish appeared healthy throughout the 8-week experimental period. The initial weight of the fish was not significantly different among experimental groups (Table 2). Final weight, percent weight gain, specific growth rate, total feed consumed and protein intake all increased, and feed conversion ratio decreased, with increasing dietary crude protein level, with significant differences among all treatment groups for all of these parameters. Protein efficiency ratio was not significantly different among the three diets with higher crude protein levels, but all were significantly higher than for the CP20 diet. Compared to initial values, body lipid content increased and water content decreased with increasing dietary protein level, with no effect on protein content (Table 3). Although there were significant differences among diets in whole-body ash and carbohydrate contents, there was no clear relationship to dietary protein level.

DISCUSSION

Growth, as expressed by absolute weight gain, percentage weight gain and specific growth rate, increased with increasing dietary protein level within the range of 20%–35% crude protein. This pattern of growth agrees with the work of Dahlgren (1979) on juvenile channel catfish; Jauncey (1982) on juvenile Mozambique tilapia, *Sarotherodon mossambicus*; Kim and Lall (2001) on juvenile haddock, *Melanogrammus aeglefinus*; Kim et al. (2002) on juvenile olive flounder, *Paralichthys oli*-

vaceus; and Martínez-Palacios et al. (2007) on juvenile Mexican silverside, *Menidia estor*. The improved growth with increasing dietary protein content in the current study was associated with increased feed consumption and protein intake by the fish. Although 35% dietary crude protein gave the best growth, the optimum protein requirement was not established because no growth plateau was found. Generally, fish reach a plateau and subsequently show decreased weight gain when dietary protein levels reach and exceed their requirements

Table 1

Component		Experimental diet				
		CP20	CP25	CP30	CP35	
Ingredient	Yellow maize ¹	48.19	38.72	29.26	19.78	
	Groundnut cake ²	19.21	28.24	37.24	46.27	
	Fish meal ³	7.69	11.29	14.90	18.51	
	Rice bran ⁴	16.06	12.90	9.75	6.59	
	Groundnut oil ⁵	5.00	5.00	5.00	5.00	
	Bone meal ⁶	2.50	2.50	2.50	2.50	
	Oyster shell ⁷	0.50	0.50	0.50	0.50	
	Vitamin premix ⁸	0.60	0.60	0.60	0.60	
	Table salt ⁹	0.25	0.25	0.25	0.25	
	TOTAL	100.00	100.00	100.00	100.00	
Proximate Composition	Crude protein	20.85	25.70	30.80	35.80	
	Crude lipid	10.85	11.20	11.60	11.90	
	Total ash	9.90	10.70	11.50	12.35	
	Crude fiber	7.40	6.50	5.70	4.80	

Composition [%] of the experimental diets containing 20%, 25%, 30%, and 35% crude protein (CP20, CP25, CP30 and CP35, respectively)

^{1–4, 6–8} Ladokun Feed Mill Ltd., Ibadan, Oyo State, Nigeria. ^{5, 9} Ficon Supermarket, Ibadan, Oyo State, Nigeria.

Table 2

Growth responses and feed utilization parameters of juvenile *Chrysichthys nigrodigitatus* fed experimental diets containing 20%, 25%, 30%, and 35% crude protein (CP20, CP25, CP30, and CP35, respectively)

Parameter	Experimental diet				
Parameter	CP20	CP25	CP30	CP35	
Initial weight [g]	21.36 ± 0.54	21.30 ± 0.43	21.38 ± 0.70	21.26 ± 0.56	
Final weight [g]	$28.21\pm0.53^{\rm a}$	$32.54\pm0.39^{\text{b}}$	$35.67\pm0.81^{\circ}$	$40.03\pm1.02^{\rm d}$	
Weight gain [%]	$32.1\pm0.9^{\rm a}$	$52.8\pm2.7^{\rm b}$	$67.0\pm9.3^{\circ}$	$88.3\pm0.2^{\text{d}}$	
Specific growth rate (SGR) [%BW/day]	$0.50\pm0.04^{\rm a}$	$0.76\pm0.03^{\text{b}}$	$0.91\pm0.06^{\circ}$	$1.13\pm0.02^{\rm d}$	
Total feed consumed (TFC) [g]	$39.06\pm0.50^{\rm a}$	$41.23\pm0.16^{\text{b}}$	$45.42\pm0.81^{\circ}$	$48.73 \pm 1.02^{\rm d}$	
Feed conversion ratio (FCR)	$5.70\pm0.06^{\rm d}$	$3.67\pm0.12^{\circ}$	$3.20\pm0.28^{\rm b}$	$2.60\pm0.12^{\mathtt{a}}$	
Protein intake (PI) [g]	$7.81\pm0.10^{\rm a}$	$10.31\pm0.42^{\rm b}$	$13.63\pm0.24^{\circ}$	$17.06\pm0.35^{\rm d}$	
Protein efficiency ratio (PER)	$0.87\pm0.01^{\text{a}}$	$1.09\pm0.04^{\rm b}$	$1.04\pm0.10^{\rm b}$	$1.10\pm0.05^{\text{b}}$	

Values are mean of 3 replicates \pm SD; values within a row having different superscript letters are significantly different at $P \le 0.05$.

(Anonymous 1993). However, the protein level of 35% is within the range reported for the optimum protein requirement for juveniles of other omnivorous fish species (Anonymous 1993), including various species of catfish (see Introduction). Generally, protein requirements vary with species, size, quality of protein, water temperature and non-protein energy sources (Craig and Helfrich 2002). In this study, the requirement for protein may have been lower if dietary lipid allowed protein sparing for growth. Further research is needed to determine the actual protein requirement of this species by increasing the protein and lipid contents of the diets.

The observation that feed conversion ratio (FCR) decreased with increasing dietary protein levels has also been reported for many other species, irrespective of culture conditions, including Mozambique tilapia (Jauncey 1982), channel catfish (Lovell 1973), African catfish (Degani et al. 1989), pike perch, *Sander lucioperca*, (see: Schulz et al. (2007), and Mexican silverside (Martínez-Palacios et al. 2007). The high values observed (2.60–5.70) may be a result of the cheap, locally available feed ingredients used for practical diet formulation; high FCR values have been reported for a number of fish species fed on practical diets using locally available feed ingredients (Khan et al. 1993)

The effects of dietary protein level on PER differ among species (Dabrowski 1979, Martínez-Palacios et al. 2007). The fact that PER was not affected by dietary protein level in the current study except with 20%CP indicates that all the other tested protein levels were used efficiently. This is in agreement with earlier studies with gilthead seabream, Sparus aurata (see: Robaina et al. 1997), European sea bass, Dicentrarchus labrax (see: Ballestrazzi et al. 1994), juvenile haddock (see: Kim et al. 2001), spotted sand bass, Paralabrax maculatofasciatus (see: Alvarez-González et al. 2001), and Mexican silverside (Martínez-Palacios et al. (2007). Other studies found that PER declined with increasing dietary protein in rabbitfish, Siganus guttatus (see: Parazo 1990), gilthead seabream (Santinha et al. 1996), olive flounder (Kim et al. 2002), and pike perch (Schulz et al. 2007). The opposite trend, that PER increases with increasing protein levels, has been observed in puffer fish *Fugu rubripes* (see: Kanazawa et al. 1980), red drum, *Sciaenops ocellatus* (see: Daniels and Robinson 1986, Serrano et al. 1992), and olive flounder (see: Kikuchi et al. 1992, Lee et al. 2000).

Initial whole-body proximate composition in the present study was higher in moisture and lower in lipid compared with the final values. These results show that gross body composition was influenced by the experimental diets, as also found for other fish species (e.g., Shearer 1994, Martínez-Palacios et al. 1996, Ng et al. 2001, Schulz et al. 2007). However, in similar studies, Lazo et al. (1998) and Alvarez-González et al. (2001) found that dietary protein level had no significant effects on proximate body composition in Florida pompano, Trachinotus carolinus, and spotted sand bass, respectively. A non-significant trend for increasing whole-body protein levels with increasing dietary protein levels, as observed in the current study, has also been reported for other species (e.g., Degani et al. 1989, Ng et al. 2001). Khan et al. (1993), however, reported that the whole-body protein content of the Asian red tail catfish increased significantly with increasing dietary protein levels up to 42% and then decreased when fish were fed with higher protein levels. Generally, there has been no consistent trend concerning the effects of dietary protein levels on fish body protein content.

Whole-body lipid levels typically decrease with increasing dietary protein levels in fish (Ng et al. 2001, Schulz et al. 2007), but the opposite effect was found in the current study. Although the same amount of groundnut oil was added to each of the diets as the lipid source, proximate analysis of the diets showed that their lipid content increased with protein content, such that the CP35 diet had 10% higher lipid content than the CP20 diet. This likely contributed to the observed increase in whole-body lipid levels with increasing dietary protein. As well, this can be attributed to the nature of this species, being a fatty fish, to deaminate and store excess dietary protein as lipid. Khan et al. (1993) reported the same effect in Asian red tail catfish. The inverse relationship between body moisture and lipid is consistent with other reports, regardless of fish species (e.g., Jauncey 1982, Ng et al. 2001).

Table 3

Whole-body chemical composition [%] of juvenile *Chrysichthys nigrodigitatus* fed experimental diets containing 20%, 25%, 30%, and 35% crude protein (CP20, CP25, CP30, and CP35, respectively)

	Initial values –	Experimental diet				
Parameter		CP20	CP25	CP30	CP35	
Crude protein	68.45 ± 0.20	68.50 ± 0.20	68.64 ± 0.19	68.84 ± 0.21	68.95 ± 0.24	
Crude lipid	$9.34\pm0.07^{\rm a}$	$9.92\pm0.20^{\rm b}$	$10.21\pm0.20^{\circ}$	$11.38\pm0.05^{\rm d}$	$12.48\pm0.09^{\text{e}}$	
Moisture	$10.01\pm0.07^{\text{e}}$	$9.31\pm0.06^{\rm d}$	$8.00\pm0.05^\circ$	$7.69\pm0.04^{\text{b}}$	$7.00\pm0.03^{\text{a}}$	
Ash	$11.13\pm0.37^{\rm c}$	$11.00\pm0.09^{\circ}$	$11.36\pm0.33^{\circ}$	$10.37\pm0.05^{\rm a}$	$10.54\pm0.23^{\text{b}}$	
Crude fiber	0.82 ± 0.002	0.64 ± 0.005	0.91 ± 0.004	0.89 ± 0.006	0.65 ± 0.003	
Carbohydrate	$0.025\pm0.006^{\rm a}$	$0.63\pm0.004^{\circ}$	$0.60\pm0.004^{\circ}$	$0.83\pm0.005^{\text{d}}$	$0.38\pm0.003^{\rm b}$	

Values are means \pm SD; values within a row having different superscript letters are significantly different at P < 0.05.

In conclusion, the use of a practical diet containing at least 35% protein is appropriate for growth and nutrient utilization of juvenile of *Chrysichthys nigrodigitatus* under the conditions of the present study.

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